## **Drill Cuttings Treatment by Bioremediation**

by

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15578

Dissertation submitted in partial fulfilment of

the requirement for the

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(Chemical Engineering)

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Universiti Teknologi PETRONAS

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### CERTIFICATION OF APPROVAL

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Approved by,

(AP DR Suzana Binti Yusuf)

## UNIVERSITI TEKNOLOGI PETRONAS

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September 2015

## CERTIFICATION OF ORIGINALITY

This is to certify that I am responsible for the work submitted in this project, that the original work is my own except as specified in the references and acknowledgements, and that the original work contained herein have not been undertaken or done by unspecified sources or persons.

## SITI HALIZAH BINTI ABU BAKAR

## ABSTRACT

This research study focuses on the removal treatment of oil residue on the drill cuttings before disposal. For this project, the method that we are interested is bioremediation process and it occurs when we mix together the sand and drilling cuts with the surfactant solutions (GraphSolve12 or GraphBioSolve). GraphSolve12 and GraphBioSolve solutions will be provided by Platinum Sdn. Bhd. as this is a joint venture project between Platinum Sdn. Bhd. and Universiti Teknologi PETRONAS. The surfactant solutions are the new solution synthesize by Platinum and the author will study the potential of GraphSolve12 and GraphBioSolve to remove oil from the drilling mud. The Total Petroleum Hydrocarbon (TPH) of the final drilling mud are compared while maintaining other parameters such as sand weight, drill cuttings weight and total amount of water and surfactant mixture. All 4 test has constant amount of water and surfactant mixture which is at 3,000 mL but different dosing percentage based on the surfactant amount such as 1ml, 10ml and 20ml for GraphSolve12 and 10mL GraphBioSolve. The Gas Chromatography-Mass Spectrometry (GC-MS) analysis show that for GraphSolve12, the best dosing is at 20% of surfactant where the result of TPH percentage shows preeminent decreasing curve of TPH percentage in the sand samples over time. It also means that the process of bioremediation is better during the 80% dosing of water for GraphSolve12. However, when we compare GraphSolve12 and GraphBioSOlve, GC-MS analysis proved that the GraphBioSolve is a better option for the bioremediation process rather than the other option because the TPH percentage achieved is lower. The TPH percentage for GraphBioSolve is 30.11% while GraphSolve12 shows 31.48% of TPH at week 10.

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## **TABLE OF CONTENTS**

CERTIFICATI	ON OF APPROVALii
ABSTRACT	iv
ACKNOWLED	GEMENTSv
CHAPTER 1: I	NTRODUCTION
1.1	Background of Study1
1.2	Problem Statement2
1.3	Objectives and Scope of Study4
CHAPTER 2: L	ITERATURE REVIEW5
CHAPTER 3: N	IETHODOLOGY/PROJECT WORK
3.1	Project Flow Chart
3.2	Gantt Chart and Key Milestones11
3.3	Preparation of Sample13
3.4	Scanning Electron Microscope (SEM) Analysis15
3.5	Gas Chromatograph-Mass Spectrometry (GC-MS) Analysis 17
	3.5.1 Extraction of Sample for Gas-Chromatography Mass-
	Spectrometry (GC-MS) Analysis (Based On TPH,
	TNRCC Method 1005, Revision 03)17
CHAPTER 4: R	ESULT AND DISCUSSION
4.1	TPH Percentage (GC-MS Analysis)19
	4.1.1 Test 1 (GraphSolve12 at 1% dosing)
	4.1.2 Test 2 (GraphSolve12 at 10% dosing)
	4.1.3 Test 3 (GraphSolve12 at 20% dosing)
	4.1.4 Test 4 (GraphBioSolve at 10% dosing)27
4.2	Surface Structure of Sand Before and After Treatment

CHAPTER 5	5: CC	<b>DNCLUSION AND RECOMMENDATION</b>	.35
5	5.1	Conclusion	.35
5	5.2	Recommendations	.36
REFERENC	ES		.37
APPENDICE	E <b>S</b>		i

## LIST OF FIGURES

Figure 1.1	Drilling Waste Management Market Size by Region from 2012 –
	2014 and the market demand on 2019 3
Figure 2.1	The changes in total resolvable hydrocarbon (A) and polynuclear
	aromatic hydrocarbon (B) concentration through the bioremediation
	period7
Figure 2.2	TPH chromatogram of the residual oil before treatment
	(WS <sub>0</sub> , WM <sub>0</sub> ) and after 12 months of bioremediation (WS <sub>12</sub> , WM <sub>12</sub> ) 8
Figure 3.1	Project Flow Chart of Research Activities10
Figure 4.1	Hydrocarbon ranges as determined by Gas Chromatography 22
Figure 4.2	The GC-MS analysis result from Sample week10 of Test 4
Figure 4.3	TPH% of GraphSolve12 at 1% dosing versus Time23
Figure 4.4	TPH% of GraphSolve12 at 10% dosing versus Time24
Figure 4.5	TPH% of GraphSolve12 at 20% dosing versus Time25
Figure 4.6	Comparison of GraphSolve12 at different dosing versus Time
Figure 4.7	TPH% of GraphBioSolve at 10% dosing versus Time27
Figure 4.8	Comparison of TPH % of GraphSolve12 and GraphBioSolve
	at 10% dosing versus Time
Figure 4.9	Surface structure of S1 native soil sand at x1.0k magnification
Figure 4.10	Surface structure of S2_T1 at x1.0k magnification
Figure 4.11	Surface structure of S3_T1 at x1.0k magnification
Figure 4.12	Surface structure of S4_T1 at x1.0k magnification
Figure 4.13	Native sand surface structure at x6.0k magnification
Figure 4.14	S2_T1 surface structure at x6.0k magnification
Figure 4.15	S3_T1 surface structure at x4.0k magnification
Figure 4.16	S4_T1 surface structure at x4.0k magnification
Figure 4.17	SEM image and corresponding EDS spectra of natural sand at
	4000x from (Tian, Gao et al. 2012)
Figure A	Details illustration on how the SEM signals are projected to obtain
	the imagei
Figure B	Step by step illustration fo the sample preparation of contaminated
	sand with surfactantii

## LIST OF TABLES

Table 3.1	Final Year Project I Gantt chart and Key Milestone	11
Table 3.2	Final Year Project II Gantt chart and Key Milestones	12
Table 3.3	GraphSolve12 Sample Preparation Data	14
Table 3.4	GraphBioSolve Sample Preparation Data	14
Table 4.1	Simple Paraffin Alkanes	20
Table 4.2	Aromatic Compounds	20
Table 4.3	Physical Parameters for TPH Aliphatic Fractions	21
Table 4.4	Physical Parameters for TPH Aromatic Fractions	21

## ABBREVIATION AND NOMENCLATURE

EPA	Environmental Protection Agency						
GC-MS	Gas Chromatography – Mass Spectrometry						
MSDS	Material Safety Data Sheets						
OBM	Oil based mud						
PPM	Part per Million						
PTFE	Polytetrafluoroethylene						
RPM	Rotation per minute						
SEM	Scanning electron microscope						
SEM-EDS	Scanning Electron Microscopy with X-ray microanalysis						
ТРН	Total petroleum hydrocarbon						
UCM	Unresolved complex mixture						
UTP	Universiti Teknologi PETRONAS						
WBC	Water based cuttings						
wt%	weight percentage						

## **CHAPTER 1**

## **INTRODUCTION**

### 1.1 Background of Study

Drill cuttings are created when a well was drilled through rock to reach an oil or gas reservoir and they can vary in size and characteristics. It happens depending on the types of drill that is used to bust the seabed during the exploration and extraction. Usually drilling mud (sand and water) will be pumped down the well to bring back the drill cuttings to the surface to keep it clean and helps to lubricate the drill bit for the oil extraction and maintain the well pressure. The disposal of the mixture (drill cuttings and drilling mud) must be properly executed to protect the aquatic habitat and reduce additional perils during pipeline maintenance. Drill cuttings can either be disposed into the sea or be taken back offshore for land disposal. However, both acts will only be allowed under the country legislation of the rig's location.

One of the most efficient drilling muds or also known as drilling fluids is oil based mud (OBM). The sticky OBM will coat the cuttings and make it hard to disperse. Even most of the OBM is removed during the process through the rig, some oil residue is still accessible on the cuttings. Upon accumulations, it will cluster below the installations and can permit lots of disturbances in future.

### **1.2 Problem Statement**

The oil and gas exploration and exploitation have become one of the main causers for the pollution in Malaysia. Even though the legislation does exist for petroleum exploration which comprise of exploration, development, production, transportation, treatment and storage, on each year, the level of pollutions keeps increasing. Currently Malaysia has six refining facilities and an average of 150 ships/day comprising of 90 cargo ships, 40 tankers and other vessels go through the Straits of Malacca (YEONG, 1990). This busy petroleum operation to meet the demands has exposed the coastal and marine habitat to contamination.

The bigger the number of oil and gas exploration, the greater the number of the drilling cuts that are available to be disposed into the sea. Over a lifetime, an oil rig can produce beyond 90,000 metric tons of drilling fluid and metal cuttings. Imagine this huge amount of waste just been thrown away into the sea and endanger our life. A recent article by Markets and Markets.com report stated that "*Recently, high growth has been noticed in drilling waste management due to increasing environmental concerns and regulatory norms imposed by the government. Major services include solids control, containment & handling, and treatment & disposal activities for safe discharge of drilling waste generated*". It shows us that in near future, by throwing drilling waste without processing it can cause you serious issues regarding environmental law obligation. Figure 1.1 shows that the drilling waste management market had increased over the years from 2012 to 2014 and the expected demand on 2019 that the world might need.



Figure 1.1 Drilling Waste Management Market Size by Region from 2012 – 2014 and the market demand on 2019

(Source: Markets and Markets.com, 2015)

As for today, water based cuttings (WBC) are directly discharged into seabed because the materials are environmentally inert and the long term effect are considered to be significant. But for OBM, the hydrocarbon disposal into the sea is a dangerous threat to the aquatic animals and human food resources. Each disposal is estimated to hold around 25,000 pounds of toxic metals and potent carcinogens such as lead, chromium, mercury, toluene, benzene, and xylene into the sea.

Due to this matters, most of the drilling cuts are being taken back to shore for land disposal. Nevertheless, this drilling cuts still contain heavy metal, hydrocarbon and other chemicals. That is why we cannot just simply throw the drilling cuts. There is numerous preferred methodology for the drilling cuts disposal alternatives. There is reduction method, recycle method and the most promising method is the waste treatment method through bioremediation before the drill cuts is being dispose. In this project, the drilling cuts will undergo bioremediation treatment and the final product will be tested for the Total Hydrocarbon Petroleum (TPH) values before disposal. This is a crucial methodology to ensure that the final product is legally disposed.

### 1.3 Objectives and Scope of Study

The main objectives of this project are:

- i. Test the performance of the GraphSolve12 and GraphBioSolve towards the bioremediation process.
- ii. Test the performance of the GraphSolve12 and GraphBioSolve towards the bioremediation process at different dosing.
- iii. Observe the difference of surface structure of sand before and after the bioremediation process.

This project will focus on the final TPH value of the drilling mud and the studied relationship between parameters. In order to achieve that, this project will focus on the outcome such as:

- i. The relation between oil contents and usage of the GraphSolve12.
- ii. The relation between oil contents and usage of the GraphBioSolve.
- iii. The relation between oil contents and time for bioremediation.
- iv. The relation between oil contents and fresh water quantity.

## **CHAPTER 2**

## LITERATURE REVIEW

According to Environmental Protection Agency (EPA), bioremediation is a treatment that uses naturally occurring organisms to break down hazardous substances into non-toxic substances. The treatment of the Alaskan shoreline of Prince Williams Sound after the oil spill of Exxon Valdez in 1989 is one common example in which bioremediation methods got public attention (Boopathy, 2000). Bioremediation technologies can be classified as *in situ* and *ex situ*. For *in situ*, it involves treating the contaminated material at the site, while *ex situ* involves the removal of the contaminated material to be treated elsewhere.

(Boopathy, 2000) said that bioremediation has numerous applications, including clean-up of groundwater, soils, lagoons, sludge and process waste streams. It has been used on a very largescale application, for instance the shoreline clean-up efforts in Prince William Sound, Alaska, after the Exxon Oil spill. However, this method does have the advantages and disadvantage depending on the microbe selection and environmental factors. The microbe is a single cell organism that are known as the oldest form of life on earth which are fungi or bacteria. The advantages and disadvantage of bioremediation are:

Advantages:

- Can be done on site
- Less expensive than other treatment methods
- Eliminates waste permanently

### Disadvantages:

- Has limitation because some chemicals are not amenable to biodegradation such as heavy metals, radionuclides and some chlorinated compound
- Sometimes, microbial metabolism of the contaminants may produce toxic metabolites.

Although bioremediation field trials were often carried out, there is insufficient information on the indigenous microbial communities that catalyse oil degradation under in situ conditions (Joel E. Kostka, 2011; Jorge Alonso-Gutierrez, 2009). The scientific factors that are affecting bioremediation are the energy sources, bioavailability, bioactivity and biochemistry. These factors have long been recognized as the parameters that influence the rate of bioremediation. Sometimes the parameter is comparably unimportant while others are crucial for a specific reaction. It shows us that certain sites may be favourably for *in situ* or *ex situ* approach.

(Atlas, 1995) has studied regarding the bioremediation to remove petroleum pollutants by seeding. He had used fertilizer in both laboratory demonstration and field demonstration and he believed that by adding a large biomass of hydrocarbon degraders, the rates of hydrocarbon biodegradation can be increased if the added cultures are able to survive and express their hydrocarbon-degradation activities in the environments to which they are added. His study has received a favourable result when both polynuclear aromatic and aliphatic hydrocarbons were biodegraded more rapidly in the fertilized than in the control shoreline sediments (Figure2.1). It is an important finding since there was concern that biodegradation might remove only some of the components of the spilled oil.



Figure 2.1 The changes in total resolvable hydrocarbon (A) and polynuclear aromatic hydrocarbon (B) concentration through the bioremediation period.

More recently (Chaillan, Chaîneau, Point, Saliot, & Oudot, 2006) research study has taken oily drill cuttings and a soil contaminated with weathered crude oils and they are evaluated by enhanced biodegradation treatment under tropical conditions in industrial scaled experiments. After 12 months of bioremediation process, the removal of hydrocarbons reached by biodegradation an extent of 60%. They also find that the residual hydrocarbons in the field treated materials were 15% - 20% further degraded when metabolic by-products resulting from biodegradation were diluted or removed under the laboratory conditions.

The result shows that the linear alkanes were not completely removed but the saturated hydrocarbons including linear, branched and cycloalkanes were degraded up to 80% (Figure 2.2). The aromatic fractions were less degraded, 38 wt% in WS and 22 wt% in WM. The aromatic unresolved complex mixture (UCM) was less assimilated than the resolved peaks confirming its resistance to microbial degradation (Chaillan et al., 2006). However, all biodegradable compounds were not

removed after 12 months as indicated by the persistence of some n-alkanes, pristane and phytane, drill fluid UCM, and bicyclic aromatics.



Figure 2.2 TPH chromatogram of the residual oil before treatment (WS<sub>0</sub>, WM<sub>0</sub>) and after 12 months of bioremediation (WS<sub>12</sub>, WM<sub>12</sub>).

For this project, the author will use *ex situ* bioremediation method as it will be done in a laboratory scale. The surfactants that will be used are GraphSolve12 and GraphBioSolve for their drilling cuts treatment. The procedures will be defined more clearly on the methodology section. The TPH percentage of the contaminated soil will be tested before and after the bioremediation treatment and samples will be taken for every two consecutive weeks and tested by using GC-MS analysis.

At the same time, the author is interested to study on the surface structure of sand used for bioremediation process. These objective can be done by using scanning electron microscope (SEM) that is used particularly for observing a fine structure of a specimen surface at high magnification. Based on (Corporation, 2009), the features of SEM are that it is applicable for all solid surfaces and can be observed in a range from low to high magnifications. SEM can allow the user to have greater focal depth than an optical microscope, allowing us to acquire a

stereoscopic image and a combination with an x-ray analyser during sample testing permits compositional analysis of a microscopic area.

The principle of how SEM works is by irradiating the sample with an electron beam in a vacuum, secondary electrons, backscattered electrons, characteristics xrays and other signals. The illustrated signals on how SEM function are given in Appendix A. In order to form an image, SEM mainly utilizes the secondary electron or backscattered electron signal. Secondary electron is produced near the sample surface, and the secondary electron image is obtained when the electrons of the fine topographical structure of sample are detected. For the backscattered electrons, it is reflected upon striking with the atom composing the sample and the electrons is dependent on the composition of atomic number, crystal orientation and others in the sample.

Before the sample is ready to be tested, it should undergo metal coating procedure. The purpose of the coating is to make the sample surface become conductive so that any charge up can be prevented. Other than that, the coating can help to increase the production rate of secondary electrons hence increasing the image formation. This metal coating will also help to prevent any damage to the sample. Generally, SEM will use gold or gold-palladium as the metal for coating by using general magnetron sputtering device. The best metal for coating is gold as it can enhance the particle to be observed at a magnification of x50, 000, x60, 000 or even higher.

## **CHAPTER 3**

## **METHODOLOGY/PROJECT WORK**

## 3.1 **Project Flow Chart**

Below is the project flow chart for this project that is recommended in order to achieve the objective.



Figure 3.1 Project Flow Chart of Research Activities

## 3.2 Gantt Chart and Key Milestones

No	Detail/Week	1	2	3	4	5	6	7	8	9	10	11	12	13	14
1	Selection of Project Topic														
2	Preliminary Research Work														
3	Submission of Extended Proposal														
4	Proposal Defence														
5	Project Work Continues														
6	Submission of Interim Draft Report														
7	Submission of Interim Report														

Table 3.1 Final Year Project I Gantt chart and Key Milestone

Process

Key Milestone

No	Detail/Week	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
1	Project Work Continues															
2	Submission of Progress Report															
3	Project Work Continues															
4	Pre-SEDEX															
5	Submission of Draft Final Report															
6	Submission of Dissertation (soft bound)															
7	Submission of Technical Paper															
8	Viva															
9	Submission of Project Dissertation (hard bound)															

 Table 3.2 Final Year Project II Gantt chart and Key Milestones

Process

Key Milestone

### **3.3 Preparation of Sample**

For this experiment, the author need to use the contaminated drilling cuts that come from an oil rig. With the help from Platinum, the drilling cuts is obtained from one of Thailand oil rig that has been posted to the Senawang, Platinum plant. In order to study the disposal effect of the drill mud to the land, some amount of sand is being mixed with the drilling cuts to act as a contaminated sand for the bioremediation process. When the contaminated sand is ready, then the surfactant is mixed together and the sample is observed. The illustration of this method is given as in Appendix B.

### Procedure:

- 1) 1kg of cutting and 4kg of soil is blended with water.
- 2) 3 litre of surfactant solution is mixed together with the sample.
- 3) The soil is submerged by approximately 1 inch under the water.
- 4) The mixture is stirred for 24 hours at 200RPM.
- 5) Pour in the flat basin and leave it for some time. The soil sample is taken periodically every 2 weeks.

For this project, GraphSolve12 will have three difference concentrations and GraphBioSolve will have only one concentration to be tested. For every test, a different ratio of water with solvent are used as stated in Table 3.3 and Table 3.4. The author has the chance to work on the sample preparation for the first sample Test 1 preparation by using 1% of GraphSolve12 as the surfactant. All the samples preparation is done by the help from Platinum Senawang plant.

Test	Cutting Cuts (kg)	Sand (kg)	Water (ml)	GraphSolve12 (ml)	GraphSolve12 Amount (%)
1	1	4	2970	30	1
2	1	4	2700	300	10
3	1	4	2400	600	20

Table 3.3 GraphSolve12 Sample Preparation Data

Table 3.4 GraphBioSolve Sample Preparation Data

Test	Cutting	Sand	Water	GraphBioSolve	GraphBioSolve
	Cuts (kg)	(kg)	(ml)	(ml)	Amount (%)
4	1	4	2970	30	10

The sample taken is transferred into a 4-ounce jar (with a Teflon<sup>TM</sup>-lined lid) and it is sent to UTP by using a standard courier services. According to (Saitas, 2001), upon received in laboratory, samples can be held at 4°C or lower if the laboratory can analyse the samples within 2 days or the samples should be placed in a maintain freezer until extracted and analysed. Because of the time constraint and the unavailability of chemicals for sample preparation for the gas chromatograph test, the samples are properly placed in the freezer once the author receive the samples. This method is based on the Total Petroleum Hydrocarbons (TNRCC) Method 1005, Revision 03, 2001 (Saitas, 2001).

### 3.4 Scanning Electron Microscope (SEM) Analysis

During this project, the author is interested to study on the difference of the sand surface structure before and after the bioremediation treatment. The experiment is conducted under the guidance of Miss Revie, student of Dr Yoshimitsu Uemura. The experiment is done by using scanning electron microscope (SEM) model TM3030-Tabletop Microscope, HITACHI. For this test, the author has chosen uncontaminated sand and 3 samples from the bioremediation process to be tested. The metal coating process is done by using a magnetron sputtering device.

### **Procedure** (Sample Preparation):

- 1) The conductive both-side tape is applied on top of a specimen stub.
- A small amount of powder is spread thinly on top of the conductive both-side tape.
- 3) Any excess powder is blow off by using a blower.
- 4) After all 4 samples are prepared, the specimen stub is place in a magnetron sputtering device for a metal coating process.
- 5) The sample is ready for SEM analysis.

## Procedure (SEM Analysis):

- 1) The instrument power is turn ON.
- 2) The PC is log in and start up the SEM software.
- The specimen stub is set on exclusive holder and the height is adjusted with "height gauge".
- 4) The AIR button on front panel of column is pressed ON to introduce air into the specimen chamber.
- 5) Gently pull out the specimen stage and set the specimen stage on the centre.
- 6) The EVAC button on the front panel of column is pressed to evacuate the specimen chamber.
- 7) The sample is ready to be observe when the evacuation of the specimen chamber is finished.
- 8) The accelerating voltage "ON" icon is clicked.
- 9) The brightness and focus is automatically being set at low magnification.
- 10) A field of interest is search and the magnification is set.
- 11) A capture box image is selected and the image is captured.
- 12) Input a file name and save the image captured.

This method is based on the Hi-Tech Instrument booklet about SEM (Corporation, 2009).

### 3.5 Gas Chromatograph–Mass Spectrometry (GC-MS) Analysis

Before we do the extraction, the sample are being dried by using a Vacuum Dryer to make sure that zero water content are available in the sample. This is because the GC-MS analysis will not give the accurate result if water still exists in the sample. For this experimental analysis, the materials and tools needed are gasoline, diesel 2-D, 99% n-pentane, 99% methanol, trifluoromethyl benzene, 1-chlorooctane, weight balance, glass jar (40mL), 10mL – 50mL volumetric flask, VOA vials with PTFE-caps, syringes, vortex shaker (optional) and Pasteur pipet.

## 3.5.1 Extraction of Sample for Gas-Chromatography Mass-Spectrometry (GC-MS) Analysis (Based On TPH, TNRCC Method 1005, Revision 03)

### **Procedure:**

- 1) The sample vial is removed from refrigeration.
- 2) Allow it to reach room temperature.
- 3) The outside of the vial is wiped with tissue.
- 4) Each vial and its contents is weighted on a loading balance and the weight is recorded to the nearest 0.01g.
- 5) The tare weight of the VOA vial is subtracted.
- 6) The resulting sample weight is recorded.
- 7) 250 ml of the Petroleum Calibration Standard is transferred into the sample using a gas-tight glass syringe. Vortex or hand shaking the sample to mix the solution for about 1 minute.
- 250 ml of the Surrogate Stock Solution is transferred into the sample using a gas-tight glass syringe. Vortex or hand shaking the sample to mix the solution for about 1 minute.
- 9) 10 ml of n-pentane is added to all samples through the septa of the vials using a 10 ml gas-tight glass syringe and vortex or hand shaking the mixture for about 1 minute. The particulate materials is let to settle within a minimum of 1 hour but can take as long as overnight.

- 10) 1-2 mL of extract sample is transferred into an auto-sampler vial using a Pasteur pipet. The auto-sampler vial is cap with a PTFE-lined cap.
- 11) Samples are ready for the GC-MS analysis by using Method 8270D.

All of the experimental work is done with the proper PPE and the MS-DS of the chemicals used are attached as the appendices.

## **CHAPTER 4**

## **RESULTS AND DISCUSSION**

### 4.1 TPH Percentage (GC-MS Analysis).

The chemical compound of sample product was analysed by using Gas Chromatography – Mass Spectroscopy (GC-MS). The TPH percentage produced at different time and dosing of surfactant is analysed for its chemical composition. Total petroleum hydrocarbon (TPH) is the measurable amount of petroleum-based hydrocarbons in an environmental matrix (Sadler R., Connell, D. 2002). Based on this study, a range of hydrocarbon from C6 – C36 are expected to be detected. According to (Sadler, R. and Connell, D. 2003) this huge range of hydrocarbon can be classified into paraffin alkenes, aliphatic and aromatics.

The lists of properties of a range of simple paraffin alkanes, which could be found in the samples are listed in Table 4.1. Then, Table 4.2 shows some corresponding physical properties for aromatic molecules that has the potency to occur in the samples. Table 4.3 and 4.4 provide some representative of physical parameters for the TPH analytical fractions based on correlations with the boiling point indices for aliphatic and aromatics.

Molecular	Name	<b>Boiling Point</b>	Melting Point	Density at
Formula		(°C)	(°C)	20°C
C <sub>6</sub> H <sub>14</sub>	n-Hexane	69	-94	0.658
C <sub>8</sub> H <sub>18</sub>	n-Octane	126	-98	0.702
$C_{10}H_{22}$	n-Decane	174	-32	0.747
$C_{12}H_{26}$	n-Dodecane	215	-12	0.768
$C_{16}H_{34}$	n-Hexadecane	287.5	18	0.775 (at mp)
$C_{20}H_{42}$	n-Eicosane	205	36.7	0.778 (at mp)
$C_{30}H_{62}$	n-Triacontane	449.7	66	0.775
C <sub>35</sub> H <sub>72</sub>	n-Pentatriacotane	490	74.6	0.781

## Table 4.1 Simple Paraffin Alkanes

## Table 4.2 Aromatic Compounds

Molecular	Name	<b>Boiling Point</b>	Melting Point (°C)	
Formula		(° <b>C</b> )		
C <sub>6</sub> H <sub>6</sub>	Benzene	80	5.5	
$C_{10}H_8$	Naphthalene	218	80.3	
C <sub>14</sub> H <sub>12</sub>	Phenanthrene	338	100.5	
$C_{18}H_{12}$	Chrysene	448	253	
$C_{20}H_{12}$	Benzo(a)pyrene	310-312	179	
C <sub>22</sub> H <sub>12</sub>	Benzo(g,h,l)perylene	542	278	

Carbon	Log S <sub>w</sub> (mg L <sup>-1</sup> )	Vapor	Henry's Law	Log K <sub>oc</sub>
Equivalent		Pressure	Constant (cm <sup>3</sup>	
Fraction		(atm)	<b>cm</b> <sup>-3</sup> )	
C <sub>5</sub> - C <sub>6</sub>	1.56	3.5 x 10 <sup>-1</sup>	47	2.9
C.>6 - C8	0.73	6.3 x 10 <sup>-2</sup>	50	3.6
C>8 - C <sub>10</sub>	-0.36	6.3 x 10 <sup>-3</sup>	55	4.5
$C_{>10} - C_{12}$	-1.46	6.3 x 10 <sup>-4</sup>	60	5.4
C>12 - C16	-3.12	7.6 x 10 <sup>-5</sup>	69	6.7
C>16 - C35	-5.6	1.1 x 10 <sup>-6</sup>	85	8.8

Table 4.3 Physical Parameters for TPH Aliphatic Fractions

Table 4.4 Physical Parameters for TPH Aromatic Fractions

Carbon	Log S <sub>w</sub> (mg L <sup>-1</sup> )	Vapor	Henry's Law	Log K <sub>oc</sub>
Equivalent		Pressure	Constant (cm <sup>3</sup>	
Fraction		(atm)	<b>cm</b> <sup>-3</sup> )	
C <sub>5</sub> - C <sub>7</sub>	2.34	1.1 x 10 <sup>-1</sup>	1.5	3.0
C <sub>&gt;7</sub> - C <sub>8</sub>	2.11	3.5 x 10 <sup>-2</sup>	8.6 x 10 <sup>-1</sup>	3.1
C <sub>&gt;8</sub> - C <sub>10</sub>	1.81	6.3 x 10 <sup>-3</sup>	3.9 x 10 <sup>-1</sup>	3.2
C <sub>&gt;10</sub> - C <sub>12</sub>	1.4	6.3 x 10 <sup>-4</sup>	1.3 x 10 <sup>-1</sup>	3.4
C <sub>&gt;12</sub> - C <sub>16</sub>	0.76	4.8 x 10 <sup>-5</sup>	2.8 x 10 <sup>-2</sup>	3.7
C <sub>&gt;16</sub> - C <sub>21</sub>	-0.19	1.1 x 10 <sup>-6</sup>	2.5 x 10 <sup>-3</sup>	4.2
C <sub>&gt;21</sub> - C <sub>35</sub>	-2.18	4.4 x 10 <sup>-10</sup>	1.7 x 10 <sup>-5</sup>	5.1

The hydrocarbons will be connected with sorbed organic matter in the soil and later the rate of the hydrocarbon sorption will differ based on the nature of the hydrocarbon (as indexed by the Koc values) and the organic matter content of the soil. Based on Table 4.3 and 4.4, the Organic Carbon-Normalized Partition Coefficient (log  $K_{oc}$ ) is depended on the water solubility [log  $S_w$  (mg L<sup>-1</sup>)].

(Sadler, R. and Connell, D., 2003) stated that the typical results that can be obtained from sand is a mixture of various hydrocarbons. Figure 4.1 below shown the GC-MS analysis range of type of hydrocarbon that is in the soil.



Figure 4.1 Hydrocarbon ranges as determined by Gas Chromatography

By referring to Figure 4.1, the GC-MS analysis reading can also provide the range hydrocarbon that the sample has. Figure 4.2 shows one of the sample GC-MS analysis result from this study and from the graph, we know that the sample contain gasoline, diesel and about 5% motor oil types of hydrocarbon ranges. From the analysis, most of the graph start after 4 minutes and end around 20 minutes later. The author can conclude that the hydrocarbon that still in the sand are mostly come from the gasoline and diesel range of hydrocarbons.



Figure 4.2 The GC-MS analysis result from Sample week10 of Test 4

## 4.1.1 Test 1 (GraphSolve12 at 1% dosing)



Figure 4.3 TPH% of GraphSolve12 at 1% dosing versus Time

From the result for Test 1 samples, the TPH line over time is irregular as it shows that the hydrocarbon is increasing and decreasing over the weeks. The TPH percentage shows good result from week 4 to week 10 as the TPH percentage decreases from 53% to 35% respectively. However, the TPH value for week 2 was so diverge and low compared to the other samples where it contains only 32% TPH value. Hence, the author can conclude that the sample at week 2 might have been continuing the bioremediation process even though the samples are properly kept inside the fridge as stated in the literature review section.



4.1.2 Test 2 (GraphSolve12 at 10% dosing)

Figure 4.4 TPH% of GraphSolve12 at 10% dosing versus Time

Figure 4.4 shows the result for Test 2 samples, the TPH line over time is irregular as does not give a linear decreasing line. The values are almost similar to Test 1 result where the TPH values are increasing and decreasing over the weeks. The TPH percentage shows more decent result from week 4 to week 10 as the TPH percentage decreases from 38% to 31% respectively. However, the TPH value (33%) for week 2 differ from the expectation where the TPH percentage is supposed

to be higher than week 4. However, the value are still higher than the TPH percentage at week 10. This shows that the error might happen during week 2 or week 4. The author can conclude that the sample at week 2 might have continued the bioremediation process even though the samples are properly kept inside the fridge or the sample at week 4 is the error. It can happen as the author is using GC-MS for this study and as we know, GC-MS is very sensitive and it has the capability to detect the hydrocarbon within the standard solution given and also other hydrocarbon outside from the standard range.



4.1.3 Test 3 (GraphSolve12 at 20% dosing)

Figure 4.5 TPH% of GraphSolve12 at 20% dosing versus Time

For Test 3 samples, the TPH value is as shown in Figure 4.5 where the line is decreasing from week 2 to week 10 which is from 45% to 30% of TPH. The decreasing line match with the author knowledge at first where the bioremediation process should be decreasing over the time. The graph shows a more stable TPH values of samples over the time and hence the author can conclude that the surfactant (GraphSolve12) give more reliable and stable results. From all the three

tests for GraphSolve12, the 20% dosing of surfactants is more reliable and effective towards the bioremediation process. The experiment should be continued so that the study can expand the scope larger to find the best surfactant amount to enhance the bioremediation process. Other than that that, the result of TPH percentage also prove to be the lowest at week 10 for Test 3.



Figure 4.6 Comparison of GraphSolve12 at different dosing versus Time

Figure 4.6 shows TPH percentage of Test 1, Test 2 and Test 3 which use GraphSolve12 as their surfactant for the bioremediation process. From the graph, the author can conclude that the best dosing of surfactant so far is during Test 3 which is 20% dosing of surfactant. The result gives the lowest TPH percentage at week 10 which is 31% and not only that, the graph shows a smooth decreasing order of the TPH percentage with respect to time. The author believes that, the amount of the surfactant also does effect the bioremediation process that happen on the microbe level.



### 4.1.4 Test 4 (GraphBioSolve at 10% dosing)

Figure 4.7 TPH% of GraphBioSolve at 10% dosing versus Time

As shown in Figure 4.7, the TPH % of Test 4 are decreasing from 31.85% at week 2 to 30.11% at week 10. However, the sample at week 8 suddenly increases and this can happen due to the condition the sample is being kept. As stated in (Method 8270D and Saitas. 2001), the sand sample taken periodically should be kept in a lined lid Teflon jar and the head gap between sample and lid should be minimize as much as possible to reduce the air gap that can affect the sand. Other

than that, the sample was not being kept immediately into the freezer after the sample was taken due to the project limitation. As the samples are being transferred from Platinum, Senawang to UTP, the journey has taken at least two days before the author received the samples and kept them into the freezer. Due to this condition, the TPH% of the samples are affected and it happen to all the samples, not particularly for test 4 samples only.

At the final week which is week 12, the TPH% show that around 30% of hydrocarbon still exist inside the sand. In order to achieve a lower TPH%, these experiment should be continued in order to obtain the time that is required to obtain low TPH% that is around 10%.

All the samples from Test 1 until Test 4 shows that the TPH value at week 10 is around 30%. According to (Chaillan, Chaîneau et al. 2006), the 12 months bioremediation process from their study als able to remove hydrocarbon to the extend of 60% which means that the TPH percentage is around 40% remain. Compared to this research study, we are able to achieve 10% lower of TPH value and also almost 9 months faster than the research study. Hence, the author can conclude that the surfactants that is used in this experiment which is GraphSolve12 and GraphBioSolve is better and will have a significant contributaion to the environment. Based on the situation, the author believe that if the new surfactant solution is used and the bioremediation process is studied until the 12<sup>th</sup> months, the result of the TPH value will decrease more to only around 1% of TPH value.



Figure 4.8 Comparison of TPH % of GraphSolve12 and GraphBioSolve at 10% dosing versus Time

Based on Figure 4.8, the comparison between the two surfactants TPH % are not significant. But if we compare based on the TPH percentage at week 10, GraphBioSolve gives a lower TPH which is 30.11%. From this findings, the author concludes that the best performance for the bioremediation process at 10% dosing is GraphBioSolve because it gives a lower TPH value compared to GraphSolve12. However, the TPH values between these two surfactant are only 1.37% difference, the experiment should be continued in order to study more on the TPH values trend that the sample can give beyond week 10. By continuing the experiment, the author believes that the final result to choose the surfactant between GraphSolve12 and GraphBioSolve will be more reliable.

### 4.2 Surface Structure of Sand Before and After Treatment.

The four samples for this study is denoted by native soil sample, S1\_T1 (Sample 1 for Test 1), S2\_T1 (Sample 2 for Test 1) and S3\_T1 (Sample 3 for Test 1). Scanning electron microscope (SEM) identifies the surface structure as illustrated in Figure 4.9, 4.10, 4.11 and 4.12.



Figure 4.9 Surface structure of S1 native soil sand at x1.0k magnification



Figure 4.10 Surface structure of S2\_T1 at x1.0k magnification



Figure 4.11 Surface structure of S3\_T1 at x1.0k magnification



Figure 4.12 Surface structure of S4\_T1 at x1.0k magnification

Figure 4.9 shows the sand surface seems to have straight and irregular steps under higher magnification (Figure4.13), the surface shows an irregular breakage blocks that looks like rough rocks. On the other hand, S2\_T1 structure is rough with angular outline like an accumulation of lots of rough rocks. The colour is noticeable that it is darker than the native soil and it proves that chemical reaction is started to affect the sand structure. From Figure 4.14, the S2\_T1 surface has a slightly weathered area (A) which mean the sand has undergone chemical weathering.



Figure 4.13 Native sand surface structure at x6.0k magnification



Figure 4.14 S2\_T1 surface structure at x6.0k magnification

S3\_T1 surface structure in Figure 4.11 show a conchoidal fracture with a straight and arcuate steps (B). The image also shows a little clearer surface as the rough rocks formation is getting lesser. Under higher magnification, the diameter of the sand crystal can be seen varies around  $10 - 40 \mu m$  (Figure 4.15). The sand particle after the bioremediation treatment is reducing its diameter and as a consequence, the quality of sand are becoming finer and better. Figure 4.12 is the surface structure of S4\_T1 shows that precipitation has occur on the sand and this happen when the area undergone intense chemical weathering (C, D). When the observation is done under higher magnification, the image in Figure 4.16 shows that the surface has a few inclined slope that is the result from the intense chemical weathering. The size of the sand particles is getting smaller and the diameter are around  $5 - 20 \mu m$ .



Figure 4.15 S3\_T1 surface structure at x4.0k magnification



Figure 4.16 S4\_T1 surface structure at x4.0k magnification

Most of the surface from Figure 4.10 - 4.12 and Figure 4.14 - 4.16, there are white colour particles that keep increasing and larger in size. (Tian et al., 2012) stated that from their finding the SEM-EDS spectrum did show the presence of metallic impurities such as Fe, Al and Ni over the figure that has lots of white particles as shown in Figure 4.17. At the same time, (Corporation, 2009) stated that in one of the example, the fibre sample observed at x4,000 magnification shows numbers of white particle that is refer to as an inorganic matter. From this study result, the white particles can be said as some inorganic materials that contain in the samples. However, the amount and types of the materials cannot be determined as more study need to be done to prove the existence. The surface structure that is

shown here can likely be a factor for the bioremediation as the transport of surfactant is affected by the porous media of the sand.



Figure 4.17 SEM image and corresponding EDS spectra of natural sand at 4000x from (Tian, Gao et al. 2012)

## **CHAPTER 5**

## **CONCLUSION AND RECOMMENDATION**

## 5.1 Conclusion

As a conclusion, this project research has successfully discovered that the best dosing for the surfactant is at 20% when mix with water. The result is based on the performance of surfactant GraphSolve12 only, but the author believe that the same result will also occur for GraphBioSolve. The experiment for GraphBioSolve is done only for one test due to the limited amount of drill cuttings. The comparison between these two surfactants also shows that GraphBioSolve is better than GraphSolve12 because it gives lower TPH percentage than GraphSolve12 at the same condition.

Overall, the bioremediation process proves to be faster and more efficient than other research findings and the author believe this surfactant will bring a big impact in the bioremediation process industry. These surfactants can help Platinum to expand their company forward and also in term of finance where they can sell the surfactant to other countries that often has major oil spills and help the environment towards a sustainable ecosystem.

### 5.2 **Recommendations**

As the research regarding bioremediation process is indefinitely wide, lots of future works can be done to further study and continue the research in other aspect of characterization of sand before, during and after the treatment. In order to commercialize the surfactants that is used in this research study, the bioremediation process should be upgraded to a bigger scale of study. The experimental works in the lab scale is only the best way to explore the process utilization of bioremediation. In future, some of the scope of study than can be explore and expanded is:

- i. Use other alternatives to obtain the TPH values from the sample other than GC-MS analysis.
- ii. Detailed study on the chemicals (surfactant) that is used in the bioremediation in advance so that the final result and the overall process is more detailed.
- iii. Run a lot more test on different dosing for both surfactants before the final decision is made for the best surfactant for bioremediation process.

### REFERENCES

- Atlas, R. M. (1995). "Bioremediation of petroleum pollutants" International Biodeterioration & Biodegradation 35(1–3): 317-327.
- Boopathy, R. (2000). "Factors limiting bioremediation technologies" Bioresource Technology 74(1): 63-67.
- Chaillan, F., et al. (2006). "Factors inhibiting bioremediation of soil contaminated with weathered oils and drill cuttings" Environmental Pollution 144(1): 255-265.
- Corporation, H. H.-T. (2009). "Let Familiarize Ourselves with the SEM." (Hitachi High-Technologies Corporation) HTD-E167P
- Joel E. Kostka, O. P., Will A. Overholt, Stefan J. Green, Gina Freyer, Andy Canion, Jonathan Delgardio, Nikita Norton, Terry C. Hazen, and Markus Huettel (2011). "Hydrocarbon-Degrading Bacteria and the Bacterial Community Response in Gulf of Mexico Beach Sands Impacted by the Deepwater Horizon Oil Spill" Applied And Environmental Microbiology. 77: 7962 - 7974.
- Jorge Alonso-Gutierrez, A. F., Joan Albaiges, Nuia Jimenez, Marc Vinas, Anna M. Solanas, and Beatriz Novoa (2009). "Bacterial communities from shoreline environments (Costa da Morte, Northwestern Spain) affected by the prestige oil spill" Appl. Environ. Microbiol. 75: 3407 - 3418.
- Method 8270D SW-846 Update V. Revision July 2014. Semivolatile Organic Compounds By Gas Chromatography/Massspectrometry. 11-13.
- Ross Sadler<sup>1</sup> and Des Connell<sup>2</sup>. (2003). "Analytical Methods For The Determination Of Total Petroleum Hydrocarbons In Soil" <sup>1</sup>Queensland Health Scientific Services, Coopers Plains, QLD. <sup>2</sup>Griffith University School of Public Health, Meadowbrook, QLD. Paper Presented At The Fifth National Workshop On The Assessment Of Site Contamination.
- Sadler, R. and Connell, D. (2002). "PAHs in the soil environment and their bioavailability in: Kookana" R, Sadler. R, Sethunathan N and Naidu, R.

Environmental Protection and Risk Assessment of Organic Contaminants. Enfield: Science Publishers, 27-43

- Saitas, J. A. (2001). "*Total Petroleum Hydrocarbons, TNRCC Method 1005*" Texas Natural Resources Conservation Commission. Revision 03.
- Tian, Y., et al. (2012). "Deposition and transport of functionalized carbon nanotubes in water-saturated sand columns" Journal of Hazardous Materials 213–214: 265-272.
- V. Ravinthar. and C.H. Yeong. (1990). "Oil Pollution in the coastal waters off Port Dickson, Straits of Malacca". Faculty of Fisheries and Marine Science, Universiti Pertanian Malaysia.

## **APPENDICES**

## **APPENDIX A**



Figure A Details illustration on how the SEM signals are projected to obtain the image image

## **APPENDIX B**



Figure B Step by step illustration of the sample preparation of contaminated sand with surfactant

## APPENDIX C



Figure C Samples preparation for GC-MS Analysis

### Example of GC-MS reading (standard at 2ppm)



## **APPENDIX E**

## Example of GC-MS reading (Test 2, Week 2)

											Area Pe	rcent Report
Da Da Acc Op San Mi: AL In In	ta Path ta File g On erator mple sc S Vial tegration tegration	: C:\( : spl: : 11 l : spl: : 8 on Para : Cher : D:\N	Jsers) 16.D Dec 20 16 Samp: ameter nStat: 4assHu	\Admin D15 : le Mu rs: au ion unter	n\De: 21:53 ltip utoin	sk 2 li s\	er: 1 .ee 1\methoo	lent	\haliz ydroca	ah	\ on.M	
Si	gnal	: TIC	C: spl	116.D	dat	a.	ms					
pea #	< R.T. min	first scan	max scan	last scan	PK TY		peak height		corr. area		corr. % max.	% of total
1 2 3 4 5	4.895 5.045 5.426 5.925 6.368	173 226 295 381 458	211 237 304 391 468	226 295 381 458 480	BV VV VV VV PV		2187216 13795119 5538991 375059 1052809	152 10 433 35 48	136432 284226 148411 217822 938120	518	2.67% 18.04% 7.60% 0.62% 0.86%	0.637% 4.305% 1.813% 0.147% 0.205%
6 7 8 9 10	6.496 6.749 6.947 7.342 7.916	480 528 560 625 691	491 535 570 639 739	528 560 625 691 766	VV VV VV VV	3 6	6101218 2069561 8014577 2280731 30548873	442 109 436 174 35	858005 073602 996349 726858 187682	51	7.77% 1.91% 7.67% 3.07% 61.73%	1.854% 0.457% 1.829% 0.731% 14.729%
11 12 13 14 15	8.183 8.407 8.558 8.914 9.339	766 809 834 861 960	786 825 851 914 988	809 834 861 960 1031	VV VV VV VV	2 3 6 3	4183208 1560236 3297056 2181865 3807278	163 46 86 191 195	432366 598590 691747 223270 674055		2.87% 0.82% 1.52% 3.35% 3.43%	0.684% 0.195% 0.363% 0.800% 0.819%
16 17 18 19 20	9.683 9.789 10.072 10.339 10.610	1031 1058 1074 1137 1201	1048 1066 1116 1162 1210	1058 1074 1137 1201 1236	VV VV VV VV VV	10 6 4 3	528926 905599 2038289 6395518 1295437	32 173 284 149	859975 792301 352571 776175 886741	7	0.85% 0.58% 3.04% 5.00% 2.63%	0.203% 0.137% 0.726% 1.192% 0.627%
21 22 23 24 25	10.881 11.046 11.154 11.268 11.451	1236 1271 1298 1313 1349	1257 1286 1305 1325 1357	1271 1298 1313 1349 1367	VV VV VV VV VV	9 3 3	2427640 3159143 2580840 10095865 2851284	172 138 80 37 108	952610 058347 452243 947777 005372	1	3.03% 2.42% 1.41% 6.66% 1.89%	0.724% 0.578% 0.337% 1.588% 0.452%
26 27 28 29 30	11.611 11.799 12.129 12.528 12.808	1367 1401 1458 1525 1585	1385 1418 1475 1545 1594	1401 1458 1525 1585 1608	VV VV VV VV	5 6 2 7	4495463 7133915 15795390 3685184 2766412	308 513 70 510 163	426187 449695 391049 101011 123935	6	5.41% 9.01% 12.35% 8.95% 2.86%	1.291% 2.149% 2.947% 2.135% 0.683%
31 32 33 34 35	12.939 13.300 13.750 14.016 14.152	1608 1668 1722 1799 1824	1617 1680 1759 1805 1829	1668 1722 1799 1824 1855	VV VV VV VV	2 2 8 8	16109259 5694335 14197843 2720629 3211703	66 463 98 198 245	498058 582272 321372 416099 133938	8	11.67% 8.13% 17.25% 3.48% 4.30%	2.784% 1.941% 4.116% 0.831% 1.026%
36 37 38 39 40	14.429 14.720 14.842 15.116 15.375	1855 1906 1947 1957 2032	1877 1928 1950 1997 2043	1906 1947 1957 2032 2057	VV VV VV VV VV	3	13058241 2389775 2503566 12454308 2493360	65 298 74 71 177	817976 966430 893853 689517 815622	8	11.55% 5.25% 1.31% 12.58% 3.12%	2.755% 1.251% 0.314% 3.001% 0.744%
41	15.513	2057	2067	2104	vv ·	7	2420973	324	886813		5.70%	1.360%

42	15.773	2104	2112	2141	vv		10424731	388182987	6.81%	1.625%
43	16.444	2169	2230	2286	vv	6	36573740	5699822705	100.00%	23.859%
44	16.812	2286	2294	2322	VV	8	2254097	217604301	3.82%	0.911%
45	17.029	2322	2332	2380	VV		12515656	432397004	7.59%	1.810%
46	17.362	2380	2390	2414	vv	4	1803285	148922862	2.61%	0.623%
47	17.584	2414	2429	2479	vv		9007072	383141104	6.72%	1.604%
48	17.920	2479	2487	2512	vv	4	1252530	105536931	1.85%	0.442%
49	18.131	2512	2524	2551	vv	-	7375342	203689349	3.57%	0.853%
50	18.450	2551	2580	2609	VV	6	935145	136209884	2.39%	0.570%
51	18,656	2609	2616	2631	vv		5533185	115828246	2.03%	0.485%
52	18.977	2662	2672	2689	vv		8280025	143337742	2.51%	0.600%
53	19.164	2689	2705	2722	vv		4083316	85467651	1.50%	0.358%
54	19.654	2780	2790	2805	vv		2503897	46056137	0.81%	0.193%
55	19.950	2805	2842	2852	VV	4	53/139	41217216	0.728	0.1/38
56	20.153	2852	2878	2909	vv	2	1022614	65794967	1.15%	0.275%
57	20.379	2909	2917	2962	vv	2	527430	37822539	0.66%	0.158%
				Cum	of	0	nroatod	20020 2300	0267617	
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hyd	rocarbon.	M Tue	Dec 1	L5 10:	38:	54	4 2015			

### **APPENDIX F**

## **MSDS of Drill Cuttings**



### MATERIAL SAFETY DATA SHEET

### 1. PRODUCT IDENTIFICATION

Product Name	Drilling Cutting, Oil Based (Fine and Coarse)
Product Description	Not Available

### 2. MANUFACTURER'S DETAILS

Manufacturer	Platinum Green Chemical Sdn. Bhd.
Address	Lot 15-19, PT 1409
	Senawang Industrial Estate
	Batu 4, Jalan Tampin
	70450 Seremban
	Negeri Sembilan
Phone	+606-6778080
Fax	+606-6770309

### 3. COMPOSITION/INFORMATION ON INGREDIENTS

Materials	Composition	CASRN	GHS Classification
C13-17 alkanes	>60%	90622-45-0	H303: Acute Toxicity (oral) Category 5, H333: Acute Toxicity (Inhalation) Category 5, H320: Eye Irritation Category 2B, H335: STOT – SE (Resp. Irr) Category 3, H336: STOT-SE (Narcosis) Category 3, H304: Aspiration Hazard Category 1,
Barium Sulfate	5 to <10%	7727-43-7	H335: STOT – SE (resp. Irr) Category 3
Calcium chloride	5 to <10%	10043-52-4	H302: Acute Toxicity (Oral) Category 4, H319: Eye Irritation Category 2A

### 4. HAZARDS IDENTIFICATION

Hazard Statement (s):	H303: May be harmful if swallowed
(-)	H333: May be harmful if inhaled
	H320: Causes eye irritation
	H335: May cause respiratory irritation
	H336: May cause drowsiness or dizziness
	H304: May be fatal if swallowed and enters airways

Pictogram	Labelling (GHS)
	• •
Signal Word	Not available
Environmental Hazards	None Identified
Precautionary statement(s):	
Prevention	P271: Use only outdoors or in a well-ventillated area
	P261: Avoid breathing dust/fume/mist/vapours/spray
	P264: Wash all exposed external body areas thoroughly after
	handling
Response	P301 + P310: IF SWALLOWED: Immediately call a POISON
	CENTER/doctor/physician/first aider
	P331: Do NOT induced vomiting.
	P304 + P312 IF INHALED: Call a POISON
	CENTER/doctor/physician/first aider if you feel unwell
	P305 + P361 + P338 : If In EYES: Rinse cautiously with water
	for several minutes. Remove contact lenses, if present and
	easy to do. Continue rinsing
	P337 + P313: If eye irritation persists, get medical
	advice/attention
Storage	P405: Store locked up
	P403 + P233: Store in a well-ventillated place. Keep container
	tightly closed.
Disposal	P501: Dispose of content/container to authorized chemical
	landfill or if organic to high temperature incineration

### 5. FIRST AID MEASURES

Eyes	Immediately flush eyes with plenty of flowing water for 10 to 15 minutes holding eyelids apart. Subsequently consult an ophthalmologist. Removal of contact lenses after an eye injury should only be undertaken by skilled personnel
Skin	Remove residues with soap and water. Change contaminated clothing. In case of skin reactions, consult a physician.
Inhalation	Provide fresh air. Seek medical treatment in case of troubles.
Ingestion	Rinse mouth and drink large quantities of water. Never give anything by mouth to an unconscious person.

	Avoid giving milk or oils Avoid giving alcohol			
	Seek medical attention.			
Indication of immediate	Any material aspirated during vomiting may produce lung			
medical attention and special	injury; hence emesis should not be induced mechanically or			
treatment needed	pharmacologically.			
	If spontaneous vomiting has occur after ingestion, the patient			
	should be monitored for difficult breathing, as adverse effect			
	of aspiration into the lungs may be delayed up to 48 hours.			

### 6. FIRE FIGHTING MEASURES

Extinguishing Media	Water spray or fog (large fire only), foam, dry extinguishing powder, carbon dioxide.
Special Hazard	Avoid contamination with oxidizing agents i.e. nitrates, oxidizing acids, chlorine bleaches, as ignition may result. In case of fire may be liberated: carbon monoxide, carbon dioxide, and acrid smoke. Mist containing combustible materials may be explosive.
Special Fire Fighting Procedure	Wear self-contained breathing apparatus. Wear suitable protective clothing.
Additional Information:	Hazchem-Code: Collect contaminated fire extinguishing water separately. Do not allow entering drains or surface water. Equipment should be thoroughly decontaminated after use.

### 7. ACCIDENTAL RELEASE MEASURES

Personal Precautions	Avoid contact with the substance.
	Provide adequate ventilation.
	Wear personal protection equipment
Environmental Precautions	Should not be released into the environment.
	Do not allow to enter into ground-water, surface water or
	drains.
	Prevent further leakage or spillage if safe to do so.
Clean-up methods-small	Absorb with liquid-binding material (e.g. sand, diatomaceous
spillage	earth, acid- or universal binding agents).
	Collect in closed containers for disposal and place in clean,
	dry, sealed container.
	Use non-sparking tools.
Additional information	Special danger of slipping by leaking/spilling product

Refer to Section 9 for additional personal protection information

Refer to Section 14 for disposal considerations.

### 8. HANDLING AND STORAGE

Handling	Avoid contact with the incompatible substance.		
	Provide adequate ventilation, and local exhaust as need.		
	Keep container sealed when not used.		
	Wear protective clothing when risk of exposure occurs.		
	Do NOT cut, drill, grind, weld or perform similar operation on or near container		
	Always wash hand with soap and water after handling.		
	Use good occupational work practice.		
	DO NOT allow material to contact exposed food or food		
	utensil		
	Work cloth should be laundered separately.		
Storage	Keep container tightly closed in a dry and well-ventilated		
	place.		
	Store away from sources of heat or ignition.		
	Store away from foodstuffs.		
Storage Class	Not available		
Specific end use(s)	Not available		

Refer to Section 7 for cleanup spillage

Refer to Section 14 for disposal considerations

### 9. EXPOSURE CONTROLS/PERSONAL PROTECTION

Avoid contact with skin and eyes.				
Change contaminated clothing.				
When using, do	NOT eat, drink	or smoke.		
Wash hands before breaks and after work.				
Have eye wash	bottle or eye ri	nse ready at wo	ork place.	
US ACGIH Tres	hold Limit Valu	es (TLV)	21	
material	material TWA STEL		Peak	
Barium	E	Not	Not	
	$1 \rightarrow 100/10^{\circ}$	27 2.5 22	\$1533 00385	
sulfate	3 mg/ m	available	available	
sulfate Emergency Lin	nits	available	available	
sulfate Emergency Lin Material	nits STEEL-1	available STTEL-2	available STEEL-3	
sulfate Emergency Lin Material Barium sulfate	nits STEEL-1 30 mg/m <sup>2</sup>	available STTEL-2 330 mg/m <sup>2</sup>	available STEEL-3 2000 mg/m <sup>,</sup>	
	Avoid contact v Change contar When using, do Wash hands be Have eye wash US ACGIH Tres material Barium	Avoid contact with skin and ey Change contaminated clothing When using, do NOT eat, drink Wash hands before breaks an Have eye wash bottle or eye ri US ACGIH Treshold Limit Value material TWA Barium 5 mg/m	Avoid contact with skin and eyes.         Change contaminated clothing.         When using, do NOT eat, drink or smoke.         Wash hands before breaks and after work.         Have eye wash bottle or eye rinse ready at work.         US ACGIH Treshold Limit Values (TLV)         material       TWA         Barium       5 mg/m.	

	according to EN 14387.
Hand/feet Protection	Neoprene gloves.
	Chemical protective gloves e.g. PVC
	Wear safety footwear or safety gumboots e.g. rubber
Eye Protection	Tightly sealed goggles according to EN 166
Body Protection	Overalls, PVC apron, eye wash unit
Additional information	Not available

### 10. PHYSICAL AND CHEMICAL

Physical State	Non slump paste	
Appearance	Liquid, does not mix with water	
Odour:	Mild characteristic odour	
Odour threshold	Not available	
Flash Point	>105 °C	
Flammability	Not available	
Explosive properties	Not explosive (method EC A14)	
Vapor Pressure	0.087 kpa	
Solubility in Water	Insoluble	
рН	Not available	
Explosive properties	None	
Auto Ignition Temperature	204 °C	
Oxidizing characteristics	Non oxidising	

### 11. STABILITY AND REACTIVITY DATA

Reactivity	No dangerous reaction known under condition of normal use.			
Chemical stability	Product is consider stable and hazardous polymerization not occur			
Possibility of hazardous reactions	Refer section 6			
Hazardous Decomposition Products	Refer section 6			
Thermal decomposition	Not available			

12. TOXICOLOGICAL INFORMATION

Animal toxicity data	Not available
Acute toxicity:	Not available
Eye Irritation	Petroleum hydrocarbon may produce pain after direct
	contact with the eyes.

	Alight, but transient disturbances of the corneal epitheli may also result.	
Inhalation	Inhalation of aerosols (mist, fumes), generated by the	
	material during the course of normal handling may be	
	damaging to the health of the individual.	
Skin Irritation	Repeated exposure may cause skin cracking, flaking or	
	drying following normal handling and use.	
	Open cuts, abraded or irritated skin should not be exposed	
	to this material.	
Ingestion	May cause abdominal pain,headache,nausea and diarrhea.	
	Large doses affect liver and kidneys.	
	May have narcotic effect	
Sensitisation	Not available	
Specific target organ toxicity	Net evelle ble	
(single exposure)	Not available	
Specific target organ toxicity	Natavailable	
(repeated exposure)	Notavanable	
Aspiration hazard	Not available	

#### 13. ECOLOGICAL INFORMATION

Aquatic toxicity:	Not available
Water Hazard Class:	Not available
Toxicity to other organisms	Not available
Biodegradation	Not available
Bioaccumulative potential	Not available
Mobility in soil	Not available

#### 14. DISPOSAL CONSIDERATION

Dispose of waste and residues in accordance with local authority requirenments.

#### 15. TRANSPORT INFORMATION

Not classified as dangerous under UN, IMO, ADR/RID, and IATA/ICAO codes

#### 16. REGULATORY INFORMATION

Cafaty, health and	
salety, nealth and	
environmental	
regulations/legislation specific	Not available
for the substance or mixture	

6

### Disclaimer.

The information provided is in good faith as a guide for handling of the product and should be treated only under the condition lay out. We cannot anticipate all conditions under which the information or our product may be used. We assume no liability or responsibility for loss or damage resulting from improper use or handling of our product from incompatible product contamination or from the failure to follow instructions, warnings and advisories in the product's Material Safety Data Sheet

## **MSDS of GraphSolve**



### MATERIAL SAFETY DATA SHEET

### 1. PRODUCT IDENTIFICATION

Product Name	Graph Solve	
Product Description	Non-ionic water based liquid blend for industrial cleaner and remediation agent.	
2. MANUFACTURER'	S DETAILS	
Manufacturer	Platinum Nanochem Sdn Bhd (Company No. 707356-X)	
Address	Lot 15-19 & PT 1409,	
	Senawang Industrial Estate,	
	Batu 4, Jalan Tampin,	
	70450 Seremban,	

+606 6778080

+606 6770309

### 3. COMPOSITION/INFORMATION ON INGREDIENTS

Materials	Composition	CASRN	GHS Classification
Proprietary blend			Acute toxicity-Oral (Category 4); Skin Corrosion/irritation
of Ethoxylated	>6004	Not	(Category 3); Eye Damage/Irritation (Category 2B); Target
Alkylphenolic	200%0	available	Organ Specific Toxicity: Single (Category 3); Target Organ
surfactants			Specific Toxicity: Repeated (Category 2)

### 4. HAZARDS IDENTIFICATION

Hazard Statement (s):	Harmful if swallowed	
	Causes mild skin irritation	
	Causes eye irritation	
	May causes respiratory irritation	
	Causes serious eye irritation	

1 of 6

Phone

Fax

Pictogram	Labelling (GHS)
Signal Word	WARNING!
Environmental Hazards	None Identified
Precautionary statement(s):	
Prevention	Use personal protective equipment as required.
	Do not handle until all safety precautions have been read and understood Obtained special instructions before use. Use only outdoors or in a well ventilated area Avoid breathing dust/fume/gas/mist/vapors/spray. Wash all exposed skin thoroughly after handling.
Response	IF swallowed: immediately call doctor/physician. Rinse
	mouth. In case of fire: use dry sand, dry chemical or alcohol resistant foam for extinction
Storage	Store locked up
Storage	Store in a well-ventilated place. Keep container tightly closed.
Disposal	Dispose of content/container in accordance with local/regional/national/international regulations.

### 5. FIRST AID MEASURES

Eyes	Immediately flush eyes with plenty of flowing water for 10 to
	15 minutes holding eyelids apart.
	Subsequently consult an ophthalmologist.
	Removal of contact lenses after an eye injury should only be
	undertaken by skilled personnel
Skin	Remove residues with soap and water
	Change contaminated clothing.
	Wash clothing before reuse.
	In case of skin reactions, consult a physician
Inhalation	Remove victim to fresh air and keep at rest in a position
	comfortable for breathing.
	If breathing is difficult, give oxygen.
	Seek medical treatment in case of troubles.
Ingestion	Do NOT induce vomiting.
	If vomiting occurs, lean patient forward or place on the left-
	side (head-down position, if possible) to maintain open
2 of 6	

	airway and prevent aspiration. Never give anything especially liquid by mouth to an unconscious person or to a person showing signs of being sleepy or with reduced awareness. Avoid giving alcohol Seek medical attention.
Indication of immediate	Any material aspirated during vomiting may produce lung
medical attention and special	injury; hence emesis should not be induced mechanically or
treatment needed	pharmacologically.

### 6. FIRE FIGHTING MEASURES

Hazardous Combustion Products	Carbon monoxide and carbon dioxide
Extinguishing Media	Small fire: Any extinguisher suitable for Class B fires, dry chemical, CO., water spray, fire-fighting foam. Large fires: Water spray, fog or fire-fighting foam. Water may be ineffective for fighting the fire, but may be used to cool fire- exposed containers.
Special Fire Fighting Procedure	Wear self-contained breathing apparatus. Wear suitable protective clothing.
Additional Information:	Use water spray to cool unopened containers. Do not allow entering drains or surface water. Equipment should be thoroughly decontaminated after use.

### 7. ACCIDENTAL RELEASE MEASURES

Personal Precautions	Avoid contact with the substance.
	Provide adequate ventilation.
	Wear personal protection equipment
Environmental Precautions	Should not be released into the environment.
	Do not allow to enter into ground-water, surface water or
	drains.
	Prevent further leakage or spillage if safe to do so.
Clean-up methods-small	Absorb with liquid-binding material (e.g. sand, diatomaceous
spillage	earth, or universal binding agents).
	Collect in closed containers for disposal and place in clean,
	dry, sealed container.
	Wash spill area thoroughly.
	Dispose of collected material according to local regulation.
Additional information	Special danger of slipping by leaking/spilling product

Refer to Section 9 for additional personal protection information

Refer to Section 14 for disposal considerations.

### 8. HANDLING AND STORAGE

Handling	Provide adequate ventilation, and local exhaust as need.
	Keep container sealed when not used.
	Wear protective clothing when risk of exposure occurs.
	Do NOT breath dust/vapor/gas.
	Always wash hand with soap and water after handling.
	Use good occupational work practice.
	No smoking.
	DO NOT allow material to contact exposed food or food
	utensil
	Work cloth should be laundered separately.
Storage	Store in cool place.
	Keep container tightly closed in a dry and well-ventilated
	place.
	Store away from sources of heat or ignition.
	Store away from foodstuffs.
	Protect container against physical damage and check
	regularly for leaks.
Storage Class	Not available
Specific end use(s)	Not available
Defer to Section 7 for algony	in anillara

Refer to Section 7 for cleanup spillage

Refer to Section 14 for disposal considerations

### 9. EXPOSURE CONTROLS/PERSONAL PROTECTION

General Precautions	Avoid contact with skin and eyes.
	Change contaminated clothing.
	When using, do NOT eat, drink or smoke.
	Wash hands before breaks and after work.
	Have eye wash bottle or eye rinse ready at work place.
Occupational Exposure Standards:	Not available
Respiratory Protection	Respiratory protection in case of aerosol or vapour formation
	Use filter type A (= against vapours of organic substances)
	according to EN 14387.
Hand/feet Protection	Neoprene gloves.
	Chemical protective gloves e.g. PVC
	Wear safety footwear or safety gumboots e.g. rubber
Eye Protection	Tightly sealed goggles according to EN 166
Body Protection	Overalls, PVC apron, eye wash unit

N/A

### 10. PHYSICAL AND CHEMICAL

Physical State	Liquid
Appearance	Red
Odour:	Wintergreen
Flash Point	Above 93 °C
Boiling Range	Not available
Explosive properties	Not available
Explosive limits	Not available
Vapor Pressure	Not available
Vapor Density	Not available
Specific Gravity	1.03 ± 0.05
Solubility in Water	Miscible
Kinematic Viscosity	Not available
рН	8.5 ± 0.5
Explosive properties	Not available
Auto Ignition Temperature	Not available
Oxidizing characteristics	Not available

### 11. STABILITY AND REACTIVITY DATA

Reactivity	Keep away from strong oxidizers and strong acids
Chemical stability	Product is consider stable and hazardous polymerization will not occur
Possibility of hazardous reactions	Not known
Hazardous Decomposition Products	Not known
Thermal decomposition	Not available

### 12. TOXICOLOGICAL INFORMATION

Animal toxicity data	Not available
Acute toxicity:	Not available
Eye Irritation	Contact with eyes may cause moderate to severe irritation.
Inhalation	Excessive exposure may cause irritation to nose, throat,
	lungs, and respiratory tract.
Skin Irritation	May cause skin irritation with prolonged or repeated contact.
	Liquid may be absorbed through the skin in toxic amounts if

	large areas of skin are exposed repeatedly.
Ingestion	There may be irritation of the throat.
	May be fatal in case of large quantity ingestion.
Sensitisation	Not available
Specific target organ toxicity	Netavoilabla
(single exposure)	
Specific target organ toxicity	Notavailable
(repeated exposure)	Not available
Aspiration hazard	Not available

### 13. ECOLOGICAL INFORMATION

Aquatic toxicity:	Not available
Toxicity to other organisms	Not available
Biodegradation	Not available
Bioaccumulative potential	Not available
Mobility in soil	Not available

#### 14. DISPOSAL CONSIDERATION

Dispose of waste and residues in accordance with local authority requirenments.

### 15. TRANSPORT INFORMATION

Not classified as dangerous under UN, IMO, ADR/RID, and IATA/ICAO codes

# 16. REGULATORY INFORMATION Not available

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